

Synthesis of 2- and 5-Diphospho-*myo*-inositol Pentakisphosphate (2- and 5-PP-InsP₅), Intracellular Mediators

Komandla Malla Reddy, K. Kishta Reddy, and J. R. Falck*

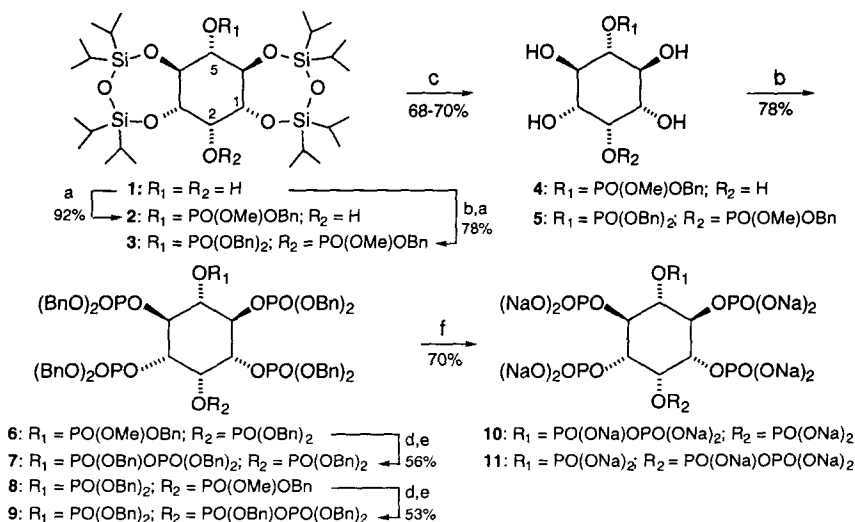
Departments of Biochemistry and Pharmacology
 University of Texas Southwestern Medical Center
 Dallas, Texas 75235-9038

Abstract: The title pyrophosphates were prepared in good overall yield from a readily available bis-disiloxanylidene derivative of *myo*-inositol. © 1997 Elsevier Science Ltd.

The intensive structural and functional studies of the *myo*-inositol cycle have led recently to the discovery of a ubiquitous family of pyrophosphoryl *myo*-inositol pentaphosphates (PP-InsP₅) and related bis-pyrophosphates.¹ Although present in most mammalian cells at 1 μM or less, the PP-InsP₅ have an unusually rapid metabolic turnover² consistent with speculation that they act as intracellular autacoids and/or as the phosphate donor for an unidentified kinase(s).³ Their levels are sensitive⁴ to changes in cellular Ca²⁺ and they display high affinity binding to several key regulatory proteins including coatomer, AP-2, and AP-3.⁵

In *Dictyostelium*, the most extensively studied organism to date, the structure of the predominate PP-InsP₅ was initially assigned as 1-PP-D-*myo*-InsP₅ or its enantiomeric 3-PP-D regioisomer.^{3a} The regiochemistry was revised to either 4- or 6-PP-D by Vogel⁶ based on 2D ¹H/³¹P NMR and was finally confirmed as the latter using synthetic standards.⁷ More recent investigations, however, have cogently demonstrated the principal PP-InsP₅ isoform produced by several mammalian cell lines is distinct from the above chiral PP-InsP₅ isomers, viz., 1-/3-/4-/6-PP-D-*myo*-InsP₅.⁸ As a consequence of the greater attention now focused on the two remaining regioisomers,⁹ we describe herein a stereocontrolled route to 5-PP-D-*myo*-InsP₅ (**10**) and 2-PP-D-*myo*-InsP₅ (**11**).

Scheme 1



Reagents and conditions: (a) **12** (1 equiv), 1*H*-tetrazole (2 equiv), CH₂Cl₂, 0°C, 3 h; *m*-CPBA, -78°C, 15 min. (b) *i*-Pr₂NP(OBn)₂, 1*H*-tetrazole, CH₂Cl₂, 0°-23°C, 4 h; *m*-CPBA, -78°C, 15 min. (c) py·HF/THF (1:2.5), 24°C, 3 h; NaHCO₃. (d) LiCN (1 equiv), DMF, 23°C, 12 h. (e) (BnO)₂POCl, Et₃N, CH₂Cl₂, 0°-23°C, 2 h. (f) Pd black/NaHCO₃, H₂ (50 psi), *t*-BuOH/H₂O (6:1), 4 h.

Access to these achiral isomers was achieved from the readily available *myo*-inositol bis-disiloxanylidene **1**, prepared according to Ozaki^{10,11} in 68% yield, as summarized in Scheme 1. Selective phosphorylation at the less hindered equatorial C(5)-alcohol in **1** via initial derivatization with benzyl methyl *N,N*-diisopropylphosphoramidite (**12**), freshly prepared from chloro-*(N,N)*-diisopropylamino)methoxyphosphine and benzyl alcohol using a modification of Caruthers' procedure,¹² followed by low temperature peracid oxidation *in situ* furnished phosphate triester **2**¹³ as an ~1:1 diastereomeric mixture. Mild de-silylation of **2** liberating pentaol **4** as expected required carefully defined conditions and was best performed with hydrogen fluoride-pyridine complex at ambient temperature. More basic/acidic reagents, e.g., *n*-Bu₄NF, HF, and CF₃CO₂H, afforded complex product mixtures. Dibenzylphosphorylation proceeded smoothly to give hexakisphosphate **6** which was advanced by specific LiCN mediated cleavage⁹ of the phosphate methyl ester. After extractive isolation, the resultant lithium salt (mp 48°C) was coupled immediately with dibenzyl chlorophosphonate as described previously⁹ to give the protected pyrophosphate **7**. Catalytic hydrogenolysis, purification by ion exchange chromatography (Q Sepharose), and bicarbonate neutralization led to the sodium salt of 5-PP-*myo*-InsP₅ (**10**).

Alternatively, prior dibenzylphosphorylation of **1** at the C(5)-alcohol and subsequent derivatization with **12** gave rise to **3** whose conversion to 2-PP-*myo*-InsP₅ (**11**) via tetraol **5**, hexaphosphate **8**, protected pyrophosphate **9**, and catalytic hydrogenolysis exactly paralleled the above sequence. The comparisons of **10** and **11** with natural material and the results of pharmacologic testing will be presented elsewhere.

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13. Spectral data for **2**: ¹H NMR (CDCl₃, 400 MHz) δ 7.30-7.45 (m, 5H), 4.85-5.25 (m, 2H), 4.26 (q, J = 12 Hz, 1H), 4.15 (dt, J = 6, 9.2 Hz, 2H), 4.01 (t, J = 2.4 Hz, 1H), 3.67 (dd, J = 2.8, 8.8 Hz, 2H), 3.55 (d, J = 11.2 Hz, 3H), 2.58 (br s, 1H), 0.80-1.25 (m, 56H). **10**: ³¹P NMR (D₂O, 162 MHz, 85% H₃PO₄ external ref.) δ 2.61 (s, 3P), 1.21-2.15 (br s, 2P), -5.25 (d, J = 14.7 Hz, 1P), -9.92 (br s, 1P). **11**: ³¹P NMR (D₂O, 162 MHz, 85% H₃PO₄ external ref.) δ 3.36 (s, 2P), 1.63 (br s, 3P), -4.90 (d, J = 16.2 Hz, 1P), -9.67 (br s, 1P).